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## **Fully automated synthesis of <sup>68</sup>Ga-labelled peptides using the IBA Synthera<sup>®</sup> and Synthera<sup>®</sup> Extension modules**

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### Introduction

The interest in <sup>68</sup>Ga-tracers has been growing strongly over the last years, mainly due to new developments in prostate cancer imaging and therapy.<sup>1,2</sup> In collaboration with IBA, we have established an automated synthesis of <sup>68</sup>Ga-labelled peptides including <sup>68</sup>Ga-DOTA-TATE, <sup>68</sup>Ga-DOTA-NOC, <sup>68</sup>Ga-PMSA and <sup>68</sup>Ga-PSMA-617 on Synthera<sup>®</sup>. The process includes elution of the generator, pre-purification of the eluate over a cation exchange cartridge,<sup>3</sup> labelling, purification and formulation of the radiotracer. The labelling, purification and formulation steps of the process would also be applicable to cyclotron produced <sup>68</sup>Ga but is not the focus of this article.

### Materials & Methods

During the tests, an IBA Synthera<sup>®</sup> and the new Synthera<sup>®</sup> Extension module were used. <sup>68</sup>Ga was obtained initially from an old iGG 100 generator in 5 mCi. The final tests were carried out in combination with a new GalliaPharm <sup>68</sup>Ge/<sup>68</sup>Ga generator loaded with 50 mCi. The generators were eluted with metal-free 0.1 N hydrochloric acid. A modified standard nucleophilic IFP<sup>™</sup> (IFP<sup>™</sup> FDG) was designed for the synthesis process. Synthera<sup>®</sup> Extension was used for the elution of the generator. The eluate was loaded on a Macherey-Nagel Chromafix PS-H<sup>+</sup> cartridge and the hydrochloric acid waste - which contains most of the <sup>68</sup>GeCl<sub>4</sub> - was transferred into the waste container. The activity was eluted from the cation exchange cartridge using a solution of 5 M sodium chloride in 0.1 N hydrochloric acid and transferred directly into the reaction vessel which was pre-loaded with precursor in 1.5 M HEPES buffer. Labelling was carried out at 95 °C for 7 minutes. The reaction mixture was taken out from the reaction vessel and the product was loaded onto a pre-conditioned Waters Sep-Pak<sup>®</sup> Light C18 cartridge. Exhaustive rinsing of the cartridge and reaction vessel with 0.9% saline removed unbound <sup>68</sup>Ga<sup>3+</sup> and HEPES buffer. The product was then eluted with a 1:1-mixture of ethanol and water and the product was dispensed into the final vial through a sterile filter. Further dilution was

performed with 0.9% saline which was also dispensed through the sterile filter. The peptides, DOTA-TATE, DOTA-NOC, PSMA-11 and PSMA-617, were synthesized in a GMP-compliant qualified area at ABX facilities. For the DOTA-peptides, stock solutions were prepared (1 mg/ml) and frozen. For PSMA-11, vials with 10 µg of precursor were used.

## Results

With 50 µg of DOTA-TATE, DOTA-NOC and PSMA-617, the final radiolabelled products were obtained in >60% uncorrected yield after 30 min of synthesis time. For PSMA-11, only 10 µg of precursor were used. The radiochemical purity was >98% in all cases. The Ph. Eur. spot test for HEPES was performed and showed HEPES < 200 µg / V with V being 12 to 14 ml. The pH of the final solution was 5 to 5.5. Ethanol content was < 10%.

## Discussion/Conclusion

We have developed a dedicated disposable IFP cassette for the IBA Synthera<sup>®</sup> and Synthera<sup>®</sup> Extension modules, which delivers all common <sup>68</sup>Ga-tracers in high yield. The use of dedicated single-use Gallium-68 IFP<sup>™</sup> allows for production of <sup>18</sup>F-FDG on the same module with no cross-contamination.

## References

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